Measurements of Natural Carbonate Rare Earth Elements in Femtogram Quantities by Inductive Coupled Plasma Sector Field Mass Spectrometry

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Supporting Information

ABSTRACT: A rapid and precise standard-bracketing method has been developed for measuring femtogram quantity rare earth element (REE) levels in natural carbonate samples by inductively coupled plasma sector field mass spectrometry that does not require chemical separation steps. A desolvation nebulization system was used to effectively reduce polyatomic interference and enhance sensitivity. REE/Ca ratios are calculated directly from the intensities of the ion beams of $^{46}$Ca, $^{139}$La, $^{140}$Ce, $^{141}$Pr, $^{146}$Nd, $^{147}$Sm, $^{153}$Eu, $^{160}$Gd, $^{159}$Tb, $^{161}$Dy, $^{163}$Ho, $^{166}$Er, $^{169}$Tm, $^{172}$Yb, and $^{175}$Lu using external matrix-matched synthetic standards to correct for instrumental ratio drifting and mass discrimination. A routine measurement time of 3 min is typical for one sample containing 20–40 ppm Ca. Replicate measurements made on natural coral and foraminiferal samples with REE/Ca ratios of 2–242 nmol/mol show that external precisions of 1.9–6.5% (2 RSD) can be achieved with only 10–1000 fg of REEs in 10–20 μg of carbonate. We show that different sources for monthly resolved coral ultratrace REE variability can be distinguished using this method. For natural slow growth-rate carbonate materials, such as sclerosponges, tufa, and speleothems, the high sample throughput, high precision, and high temporal resolution REE records that can be produced with this procedure have the potential to provide valuable time-series records to advance our understanding of paleoclimatic and paleoenvironmental dynamics on different time scales.

Since the 1980s, rare earth elements (REEs) have emerged as important tracers for diverse applications of the earth sciences, including evolution of lithospheric reservoirs, modern and past environmental changes, and marine geochemistry and ocean circulation. Records of REEs in natural carbonate materials, such as corals, foraminifera, and speleothems, have been analyzed for understanding contemporary and late-Quaternary climatic and environmental changes. However, overall applicability has been sharply limited due to the low REE abundances, 1–100s nmol/mol, in most natural samples and challenging analytical difficulties. Different analytical methods, such as neutron activation analysis, inductively coupled plasma atomic emission spectrometry, inductively coupled plasma quadrupole mass spectrometry (ICP–QMS), isotope dilution thermal ionization MS (ID–TIMS), cathodoluminescence, and laser ablation ICP–MS (LA-ICP–MS), have been employed for carbonate REE determinations. ID–TIMS can deliver good precision of 0.1–3% (2 relative standard deviations, 2 RSD); however, labor-intensive sample preparation processes limit the rate of analytical measurements. With improvements in instrumentation, including
high sensitivity, low detection limit, and rapid multielement analysis in the past decades. ICP—MS provides the ability to analyze carbonate REEs with a precision of 6—20%\textsuperscript{,15,16} For both ID—TMS and ICP—MS, time-consuming column chromatography is usually required to separate REEs from a matrix, and this step ultimately hinders sample throughput. Simple ICP—MS methods using enriched isotopes and elemental internal standards without chemical separation steps give a 2 RSD of 10—100% for nanomoles per mole level REE analyses\textsuperscript{,12,18} Modern LA-ICP—MS methods can offer a direct and fast measurement of micrometer-resolution carbonate REE concentrations\textsuperscript{,9} however, low REE abundances in corals sharply limit the 2 RSD precision to 26—36% using this procedure.\textsuperscript{9}

In this study, we established protocols to directly measure carbonate REE abundances by ICP-sector field (SF)—MS with a 2 RSD reproducibility of 1.9—6.5% for 10—20 μg carbonate samples after their dissolution. We carefully addressed factors affecting the high-precision determination of REE/Ca ratios: (1) spectral interferences\textsuperscript{,19,20} (2) mass discrimination and ratio drift\textsuperscript{,21,22} and (3) chemical matrix effects.\textsuperscript{21,23} Examples include modern Porites corals, planktonic foraminifera Globorotalia menardii, and benthic foraminifera Cibicides wuellerstorfi.

# EXPERIMENTAL SECTION

Reagents, Standards, and Samples. Preparation of standards and samples was performed in a class-10 000 geochemical clean room with class-100 benches in the High-Precision Mass Spectrometry and Environment Change Laboratory (HSFPEC), National Taiwan University. Water was purified using an ultrapure water tandem system with Millipore Milli-Q Academic and Milli-Q Element. PTFE and polyethylene vials, bottles, and beakers were cleaned by boiling with 3 N guaranteed reagent Milli-Q Element II ICP Element II ICP spray chamber and desolvator were set at 110 and 160 °C respectively. This system provided a 5—10-fold enhancement in sensitivity and dramatically reduced the polyatomic interferences from hydrides and oxides.\textsuperscript{53} Overall sensitivity was 1.5—2.0 × 10^6 cps ppb\textsuperscript{-1}. The ASX-100 Micro Autosampler (CETAC Technologies, NE) was utilized for automatic sequence measurements. A single secondary electron multiplier in peak-hopping mode was used to measure the ion beams of 46Ca, 138Ba, 139La, 140Ce, 141Pr, 146Nd, 147Sm, 153Eu, 154Tb, 156Gd, 157 Dy, 158Ho, 160Er, 169Tm, 172Yb, and 175Lu. Ion beam intensities of 46Ca and 138Ba were measured in analog mode and REEs were measured in ion-counting mode. Cross-calibration between analog and ion-counting modes was performed using a 46Ca ion beam of 0.8—1.5 × 10^6 cps before running samples, and instrumental drift was calibrated by measuring bracketed standard solutions between samples. Magnetic-scan (B-scan) mode was used for peak jumping between masses 46, 138, and 159, and electrostatic-scan (E-scan) mode for masses of 139—153 and 159—175. For each measurement, sample solution uptake lasted for 190 s, followed by a 110-s washout step with 5% HNO₃. Every four samples were bracketed with one standard. All REE/Ca ratios were calculated directly from ratios of ion beam intensities using external matrix-matched standards to correct for instrumental mass discrimination and ratio drifting. Data were calculated in an off-line data reduction process, modified from Shen et al.\textsuperscript{25} All errors given are two standard deviations (2σ) or 2 RSD unless otherwise noted. Detailed instrumental settings and data acquisition methods are summarized in Supporting Information Tables S2 and S3.

## RESULTS AND DISCUSSION

Blanks and Spectral Interferences. The procedural blank (PB), including chemical blank and spectral interferences, is
<2 × 10^3 cps at 46 atomic mass unit (amu) and <70 cps at 139–175 amu. The PB/signal ratio is 0.1–0.2% for 46Ca and <0.14% for all REE isotopes of interest in CarbREE-I standard solution with 20 ppm Ca. For natural carbonates with <10 s nmol/mol REE/Ca, the PB/signal ratio is 2–3% for 153Eu, 159Tb, 160Gd, and 163Dy, and <1% for other measured REE isotopes. There are two types of instrumental blank in our ICP–SF-MS. One is a residual signal from previous samples. After the uptake of 5% HNO3 for 70 s, the calcium and REE blanks were reduced to <0.03% of the original signal. These blanks were subtracted from measured ion beams to earn net intensities. Another type of memory interference is a "ghost" ion beam due to a significant contamination after running high concentration calcium samples for 1–2 h. These unexpected abrupt spiky signals were filtered in an off-line data reduction process.

Serious isobaric interference of oxide and hydroxide forms of Ba on Eu was reported for wet introduction systems. The BaO/Ba and BaOH/Ba ratios are only 41.9 × 10^{-6} and 2.9 × 10^{-6}, respectively, using the desolvation introduction system (Figure S1). Ba/Ca ratios range from ∼1 to 10 μmol/mol in marine carbonates of corals and foraminifera. The 137BaO signal is <10 cps and the 136BaOH signal is <1 cps at 153 amu. The correspondent isobaric interference for 153Eu is <2.5% for samples with low REE levels of 1–10 nmol/mol. The 137BaO and 136BaOH can cause a high isobaric interference, ∼20% on the 153Eu ion beam, for carbonates with a Ba/Ca level >100 μmol/mol and Eu/Ca <10 nmol/mol. Hence, we measured the 136Ba+ ion beam and corrected the isobaric interference using an intensity ratio of 10.7 × 10^{-6} [(137BaO+ + 136BaOH+)/138Ba+] for all carbonates. 140Ce16O+ is an isobaric interference for 156Gd+ measurement. To avoid this spectral interference, 160Gd+ was selected to determine Gd content.

Mass Discrimination and Isotopic Ratio Drifting. Increased ion transmission with mass causes deviations of measured ratios from true ratios, and this mass discrimination (or mass bias) affects the analytical precision and accuracy of ICP–MS measurements. Low-frequency drift of measured isotopic ratios was also observed (Figure 1A), mainly as a result of an analog measure mode used for 46Ca+ and 138Ba+, deposition from high-calcium samples, various plasma conditions, and drift in the electronics. One typical example of low-frequency drift of measured La/Ca ratios is shown in Figure 1B. La/Ca ratios can vary 20% in 2 h. To correct for these types of biases, the CarbREE-I standard solution was measured after every 4 samples. This allowed for a linear-interpolation correction with a bracketing standard. With the correction, a short-term external 2 RSD precision of 0.8% for La/Ca was achieved (Figure 1B).

Matrix Effects. We evaluated the acid effect, which is one type of matrix effect observed in ICP–MS using wet introduction systems with ion beam intensities of different elements. As an
example, the La/Ca ratios of a 20 ppm-Ca CarbREE-I standard solution in different strengths of HNO₃ ranging from 1 to 6% are plotted in Figure 2. The measured La/Ca ratios for 6% HNO₃ are slightly higher than the other values, showing a minor acid effect. The external errors of 1.6—2.4% for 1—3% HNO₃ are 1.5—2 times larger than the internal errors of 1.1—1.3%, probably related to the formation of nitrate particulates during the desolvation process. The small external errors for high HNO₃ levels of 4—5% are comparable to the internal precisions. A compensative 5% HNO₃ with good reproducible data was used for routine measurements.

Previous studies²²,³⁴ have shown a matrix effect of Ca concentration on accurate elemental ratio determinations. This effect was addressed by measuring REE/Ca ratios in a range of diluted standard solutions with Ca levels of 5—50 ppm. Figure 3 shows the Ca matrix effect on La/Ca and Yb/Ca. For [Ca] of 5—20 ppm, the measured La/Ca ratio decreases with a slope of \( \sim 17 \text{ La/Ca (nmol/mol)/[Ca] (ppm)} \). For [Ca] ≥ 20 ppm, the Ca matrix effect is trivial (Figure 3). A similar Ca matrix effect on Yb/Ca measurement with decreasing trends is observed. A 2—4% change in Yb/Ca between 20 and 40 ppm [Ca] is observed. These observed decreasing REE/Ca trends are likely attributed to space charge effects.³⁵ REE ions could be strongly defocused within the skimmer with high calcium ion intensities. To attenuate the [Ca] matrix effect, [Ca] in both sample and standard aliquots was adjusted to the same level (within 10%) between 20 and 40 ppm prior to measurement.

One disadvantage for this fast carbonate REE measurement method is inevitable Ca deposition on sampling cones. When running samples with 40 ppm Ca concentration, the intensities of the ion beams decrease by a typical rate of \( \sim 10\% / \text{h} \). The sampling cones should be cleaned manually every day. We suggest analyzing samples with a low Ca concentration of 20 ppm if they have relatively high REE/Ca ratios of tens to hundreds of nanomoles per mole. The daily cleaning step for the sampling cones will not be necessary.

Accuracy. REE concentration uncertainties in the commercially available REE solution standard are 1.0%, based on the reports given by High-Purity Standards (Charleston, SC). The use of superpure Sigma-Aldrich calcium carbonate powder contributes 13.6% (±3.0%) for La, 35.5% (±3.0%) for Ce, and

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**Figure 4.** Percent offset of measured REE/Ca ratios with long-term 2 RSD error bars (Table S4) from theoretical values in the CoralM-REE standard solution. The shaded area denotes the uncertainty of REE/Ca in the standard.

**Figure 5.** Comparison of measured REE/Ca ratios of a benthic foraminifer, *C. wuellerstorfi*, from marine core MW91-9 GGC-15 by our ICP–SF-MS and ICP–QMS²²,²⁸ at the University of Cambridge.

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**Figure 6.** Replicate measurements of La/Ca (circles) and Yb/Ca (diamonds) with an internal error of measured REE/Ca ratios in (A) CarbREE-I, (B) coral ST0506, and (C) foraminifer FORAM-GM. External errors are given with a dashed line and values in gray.
<0.3% for all other REEs in the gravimetrically prepared CarbREE-I solution standard. The uncertainties of true REE concentrations in this CarbREE-I standard, including impurities from the calcium carbonate powder, are estimated to be ±1.5% for La, ±3.2% for Ce, and ±1.0% for Pr, Nd, Sm, Eu, Gd, Tb, Dy, Ho, Er, Tm, Yb, and Lu. Measured low CarbREE-II REE/Ca ratios determined with CarbREE-I are insignificantly different from gravimetric values (Table S1), suggesting a good linearity of intensity signals.

The differences between determined REE/Ca ratios and gravimetrically calculated values in the admixed coral standard solution, CoralM-REE, are all less than the long-term uncertainties of 2−6% for coral carbonate (Figure 4). This indicates there is no significant matrix effect to quantify REE contents in natural carbonates by using the in-house standards. An interlaboratory comparison for a benthic foraminifer C. wuellerstorfi sample from core GGC-15 shows that our ICP−QMS results replicate measurements made by ICP−QMS at the University of Cambridge (Figure 5), with the exception of Ce/Ca. This sole discrepancy (13±4% for Ce/Ca) is attributed to slightly heterogeneous Ce contents between the two subsamples that were used in the two laboratories. The above-mentioned experiments indicate our ICP−QMS methods can accurately be applied to natural carbonates without a chemistry separation process.

**Precision, and Detection Limit.** Analytical reproducibility over the course of 6 months (Dec 2008−May 2009) determined by repeatedly analyzing the CarbREE-I standard solution is plotted in Figure 6A. Internal 2 RSD precision of REE/Ca ratios ranges from 0.4 to 1.4%. Replicate measurements show that long-term reproducibility ranges from 0.8 to 1.7%, 1.5−2 times that of internal precision. An external uncertainty, for example, is 0.9% for La/Ca and 1.1% for Yb/Ca, respectively (Figure 6A).

Two-month (July−August 2010) REE/Ca data of coral ST0506 (2−242 nmol/mol) and foraminifer FORAM-GM (3−157 nmol/mol) are given in Table S4, and La/Ca and Yb/Ca are also illustrated in Figure 6B,C. Internal 2 RSD precision ranges from 1% to 2% for high (tens to hundreds of nanomoles per mol) and low (<10 nmol/mol) REE levels, respectively. External uncertainties of 1.9−6.5% are 2−4 times larger than internal ones.

Large external errors of 5−6% are observed for Eu, Gd, and Dy in coral ST0506 (Table S4). Relatively high 2σ external deviations of 5−8% and positive offsets of 3−4% are also observed for Gd and Dy in the reference sample CoralM-REE (Figure 4). The high deviations and offsets are likely caused by polyatomic interferences from trace elements or organic matter or complexes or both, indicating the matrix complexity of natural carbonates.

On the basis of the definition of the detection limit, 3×σB, where σB is the standard deviation of the procedural background of 10−70 cps on ICP−QMS, the calculated detection limits of carbonate REE/Ca ratios range from 0.1 to 0.7 nmol/mol. The Eu detection limit varies with Ba content on ICP−MS (see section “Blanks and Spectral Interferences”). The isobaric spectral background should accurately be applied to natural carbonates without a chemistry separation process.

A striking feature of the monthly resolved REE time series is a clear seasonal cycle, from low levels of 40−80 ppb in summer to high values of 120−140 ppb in winter/early spring (Figure 7). Individual REE/Ca ratios, even as low as 0.3−1 nmol/mol for Eu, Tb, Lu, and Ym, also show coherent seasonal variability, along with total REEs (Figure 7). High coastal seawater REE levels are usually delivered by terrestrial runoff during the rainy season.8,9 In contrast, the high WZI-1 coral REE levels are observed in the dry winter/early spring season (Figures 7, S3).

WZI, a small island with an area of 25 km², is isolated from mainland China, well-protected with a dense vegetation cover.

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**Figure 7.** Monthly resolved time series for the 4-year period (AD 2002−2005) of (A) La/Ca, Nd/Ca, and Ce/Ca; (B) Gd/Ca, Sm/Ca, and Pr/Ca; (C) Ho/Ca, Eu/Ca, and Tb/Ca; (D) Dy/Ca, Er/Ca, and Yb/Ca; (E) Lu/Ca and Tm/Ca; (F) total REEs; and (G) Sr/Ca for the Porites coral, WZI-1, and (H) monthly sea surface temperature (SST) from a WZI meteorological station. Coral REE time series were calendared by aligning coral Sr/Ca maximums with SST minimums. The error bars of REE data are smaller than the symbol size.
and lack of riverine sediment input and local anthropogenic inputs. The most probable source for high REE values is from aeolian dust transported by winter monsoon winds. East Asia is a primary source for natural and anthropogenic dust aerosols in the Northern Hemisphere. The strong and dry winter monsoon primary source for natural and anthropogenic dust aerosols in the Northern Hemisphere.38 The strong and dry winter monsoon not only brings cold air masses but also transports large amounts of dust (average deposition: 2–5 g/m²/year) to eastern and southern China and beyond,38,39 recorded as high REE levels in WZI corals. This high-resolution coral REE fingerprints could be new proxies for East Asian winter monsoon.

Summary and Conclusions. After carefully addressing issues, including spectral interferences, blank and memory effects, ratio drifting and mass discrimination, and matrix effects, a precise procedure for measuring carbonate REEs on ICP–SF-MS has been developed. A high instrumental sample throughput of 8–10 samples/h can be achieved routinely. This high-sensitivity method with good external reproducibility (1.9–6.5%) requires only 10–1000 fg of REEs in 10–20 μg of carbonate samples. Compared with labor-intensive TIMS methods,6,10 only simple cleaning and dissolution steps are involved, and no time-consuming column separation is required in this procedure. The analytical precision is superior to that of previous ICP–MS16,18 and LA-ICP–MS methods.8,9 A clear seasonal cycle of low WZI coral REE/ Ca ratios <1 nmol/mol shown by our fast ICP–SF-MS method is difficult to achieve using any other current ICP–MS technique. The examples described here demonstrate the applicability of high-precision monthly coral REEs as proxies for detailed dust history and monsoon evolution. In addition to corals and foraminifera, our method can also be applied to diverse carbonates, such as spherulites, tuñá, shells, and speleothems. Hence, the carbonate REE-related studies of climate change,12 environmental research,8,13 and paleoceanography11 can benefit from the improvements in precision, sample size, and sample throughput inherent in this new ICP–SF-MS method.

ASSOCIATED CONTENT

Supporting Information. Additional information as noted in the text. This material is available free of charge via the Internet at http://pubs.acs.org.

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